Quantification of Beta-Hydroxybutyric acid and Tryptophan in plasma as metabolic biomarkers of cancer using the LDTD-MS/MS technique



OVERVIEW

Purpose

• Automated extraction process and LDTD-MS/MS quantification of Tryptophan and Beta-Hydroxybutyric acid in plasma

Method

- Automated Assisted Liquid-Liquid Extraction
- Samples dried and analyzed by LDTD-MS/MS

Quantification

- Linearity: $r^2 > 0.99$ over the calibration range
- Between-run accuracy, values between 86,0% and 107,6% were obtained and the precision results were lower than 15% CV.
- Samples analyzed with a runtime of 8 seconds using LDTD-MS/MS technique

INTRODUCTION

Metabolomics has the potential to map early biochemical changes in cancer cells and hence provides an opportunity to quantify cancer biomarkers in blood that can trigger earlier diagnosis. Imbalances in tryptophan (TRP) metabolism have been linked to cancer-related immune escape and implicated in several cancers, including lung cancer. Accumulated metabolites of β hydroxybutyric acid (β-HBA) from higher energy production by cancer cells is considered to be one of the hallmarks of cancer development. Therefore, the rapid and accurate determination of Tryptophan and β -HBA concentration in plasma would be useful for the development of an early cancer diagnosis tool.

For this project, an automated extraction method is developed. TRP and β-HBA drugs in plasma are extracted and quantification using Laser Diode Thermal Desorption and tandem mass spectrometry (LDTD-MS/MS) is chosen as a fast-analytical technique.

LUXON Ionization Source:

The Luxon Ion Source (Figure 1) is the second-generation sample introduction and ionization source based on the LDTD technology for mass spectrometry. The Luxon Ion Source uses a Fiber-Coupled Laser Diode (Figure 2) to obtain unmatchable thermal uniformity giving more precision, accuracy and speed. The process begins with dry samples which are rapidly evaporated using indirect heat. The thermally desorbed neutral molecules are carried into a corona discharge region. Highefficiency protonation and strong resistance to ionic suppression characterize this type of ionization and is the result of the absence of solvent and mobile phase. This thermal desorption process yields high-intensity molecular ion signal in less than 1 second sample-to-sample and allows working with very small volumes.



Figure 1 Luxon Ion Source



Figure 2 Schematic of the Luxon Ion Source

Anatole Pelletier ⁽¹⁾, Jean-François Haince ⁽¹⁾, Jonah Randia ⁽¹⁾, Rashid Ahmed Bux ⁽¹⁾, Jean Lacoursière ⁽²⁾ and Serge Auger ⁽²⁾ ⁽¹⁾ BioMark Diagnostic Solutions Inc., Québec, QC, Canada ⁽²⁾ Phytronix Technologies, Québec, QC, CANADA;

METHOD

Automated Assisted Liquid-Liquid extraction

complete dryness before analysis.

Instrumentation

Luxon Parameters

0%

• Laser power pattern:

65% in 6 sec

• Carrier gas flow: 6 L/min (Air)

- Hold 2 seconds

- Increase laser power to

- Decrease laser power to

45 μ L of plasma are mixed with 45 μ L of the internal

standard solution (TRP-d5 and β -HBA-d4, 30 μ g/ml and 20

 μ g/ml in methanol:water 25:75). 5 μ L are transferred into

a deep-well extraction plate on a vortexer system and

mixed with 250 µL of methanol:water (1:1). Finally, 5 µL of

the extraction and then 5 μ L of desorption solution were

spotted onto LazWell[™] 96 plates and evaporated to

• Ion source: Phytronix, Luxon S-960 Ion Source

• Mass spectrometer: Sciex, Q-Trap System 6500+

Validation test

Linearity

RESULTS

Quality Control (QC) plasma samples were prepared using the SeraCon[™] I Negative Diluent and the endogenic concentration values were determined by extrapolation of the calibration curves, a correction of the standards solutions was then applied and used as a target value. Repeated extractions were deposited onto a LazWell[™] plate and dried before analysis. The peak area against the internal standard (IS) ratio was used to normalize the signal.

linearity test, the following acceptance criteria was used:

- Linear regression (r) must be ≥ 0.995

Table 3 shows the inter-day correlation coefficients for TRP and β -HBA. Values greater than 0.995 are obtained. **Figure 4** shows a typical calibration curve result for β -HBA.

The calibration curves were plotted using the peak area ratio and the nominal concentration of standards. For the

Table 1 MS parameters Compound β-ΗΒΑ TRP APCI mode Curtain 20 20 CAD 30 25 Dwell Time

Compound	Q1 (Da)	Q3 (Da)	CE (V)
TRP	205	118	35
TRP-d5	210	122	35
β-ΗΒΑ	103	59	-15
β-HBA-d4	107	59	-15



Figure 3 Azeo: Automated extraction system

RESULTS



Figure 4 β-HBA calibration curve

Stability

Wet stability of Sample Extracts:

Following the extraction, sample extracts are kept at 4°C in closed containers. After 1 day, sample extracts are spotted on a LazWell[™] plate, dried and analyzed. Precision and accuracy of QC samples are reported in **Table 5.** All the results are within the acceptable criteria range for 1 day at 4°C.

Table 2 MRM transitions parameters

Dry stability of Samples spotted in LazWell:

Extracted samples are spotted onto a LazWell[™] plate, dried and kept at room temperature for 2 hour before analysis. The precision and accuracy results of QC samples are reported in **Table 5.** All the results are within the acceptable criteria range for 2 hour at room temperature.

Table 5 Wet and Dry stability of TRP and β -HBA

Parameters	Dry stability (2 hour / RT)			ability (2 hour / RT) Wet stability (1 day / 4°C)				
molecule	TRI	D	β-H	IBA	TR	Р	β-H	IBA
QC	QC1	QC2	QC1	QC2	QC1	QC2	QC1	QC2
Conc. (µg/mL)	17,8	62,8	17,0	62,0	15,8	60,8	16,2	61,2
Ν	3	3	3	3	3	3	3	3
Mean (µg/mL)	15,9	60,4	16,4	62,7	17,2	55 <i>,</i> 43	16,5	54,4
%CV	4,18	0,34	3,91	0,82	3,44	2,35	2,47	2,86
%Target	93,5	96,1	96,7	101,3	108,7	91,1	101,8	88,8

CONCLUSION

- High-throughput analysis using LDTD-MS/MS
- Linearity, accuracy, precision and stability within the acceptance criteria
- Sample-to-sample analysis of 8 seconds
- System can be customized for molecular detection of cancer

Phytronx

Table 3 Inter-day calibration curve correlation

	TRP	β-ΗΒΑ
Curve 1	0,9986	0,9998
Curve 2	0,9979	0,9996
Curve 3	0,9986	0,9987
Curve 4	0,9978	0,9967
Curve 5	0,9979	0,9996

Precision and Accuracy

For the accuracy and precision evaluation, the following acceptance criteria were used:

- Each concentration must not exceed 20% CV
- Each concentration must be within ±3 SD of the target value

For the inter-run precision and accuracy experiment, each standard was analyzed in triplicate, on five different days. Tables 4A and 4B show the interrun precision and accuracy results for TRP and β -HBA . The obtained %CV was below 10% and the accuracy was within ±3 SD of the target value.

Table 4A	Inter-run	precision and	d accuracy for	or TRP
		P		

TRP	QC1	QC2
Conc (ng/ml)	15,2	60,2
Ν	15	15
Mean (ng/mL)	16,4	51,8
SD	1,4	4,0
%CV	8,6	7,7
%Target	107,6	86,0

Table 4B Inter-run precision and accuracy for β -HBA

β-ΗΒΑ	QC1	QC2
Conc (ng/ml)	17,6	62,6
Ν	15	15
Mean (ng/mL)	17,5	60,9
SD	1,6	3,4
%CV	9,2	6,5
%Target	101,1	97,6

• Efficient Automated Assisted Liquid-Liquid Extraction is used to extract Tryptophan and β-Hydroxybutyric acid in plasma

